# Detection of Gastrin in Formalin-Fixed, Paraffin-Embedded Rat Tissue

### **Reagent and Antibody Information**

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
Carezyme II (Pepsin)
DAB Chromogen
Hematoxylin

Blocking Solution: Dakocytomation Protein Block Serum-Free Ready-To-Use

Dakocytomation Corporation Carpinteria, CA 93013 www.dako.com 1-800-235-5763 Code No. X0909

## Avidin / Biotin Blocking Kit

Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Rabbit Polyclonal Anti-Human Gastrin

Dakocytomation Corporation Carpinteria, CA 93013 www.dako.com 1-800-235-5763 Code No. A0568

Negative Control Serum: Rabbit Immunoglobulin Fraction (Solid-Phase Adsorbed)

Dakocytomation Corporation Carpinteria, CA 93013 www.dako.com 1-800-235-5763 Catalog # X0936

Staining Kit: LSAB+ System-HRP

Dakocytomation Corporation Carpinteria, CA 93013 www.dako.com 1-800-235-5763 Code No. K0690

**Note**: This kit includes reagents needed for the secondary antibody (link) and label complex.

### **Staining Procedure**

Positive Control Tissue: Gastrointestinal Tract

Stain Localization: Cytoplasmic

1. Deparaffinize and hydrate slides through the following solutions:

Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.
- 4. <u>Proteolytic-Induced Epitope Retrieval Using Pepsin</u>

Prepare a 1:3 dilution of the Carezyme II: Pepsin reagent (1 part pepsin and 2 parts distilled water) Preheat slides to 37°C in 1X wash buffer.

Incubate the slides in the pepsin solution for 90 seconds at 37°C.

Rinse the slide in distilled water for 1 minute to stop the enzymatic reaction.

- 5. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.
- Block with the Dako protein-blocking reagent for 10 minutes at room temperature.

  Lot #\_\_\_\_\_ Exp. Date\_\_\_\_\_

  DO NOT RINSE THE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.

# 7. Avidin / Biotin Blocking Kit

Lot #\_\_\_\_ Exp. Date\_\_\_\_ New Kit: yes / no Apply avidin block for 15 minutes at room temperature.

Quick rinse in 1X wash buffer.

Apply biotin block for 15 minutes at room temperature.

DO NOT RINSE SLIDES WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY. ONLY WIPE EXCESS BLOCK.

8. Apply	the primary antibody	at a 1:300 dilution	n. Incubate for 30	) minutes at room	temperature.
Lot #	Exp	. Date			

For negative control slides, dilute normal rabbit IgG so that it's IgG protein concentration matches that of the primary antibody (if necessary). Then make a 1:300 dilution. If the concentrations can't be matched using this method, the dilution for the negative reagent may need to be adjusted. Apply the negative and incubate for 30 minutes at room temperature.

Lot #\_\_\_\_\_ Exp. Date \_\_\_\_\_

9. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.

LSAB+ Kit Lot #	Exp Date				
10. Apply the Link (yellow bottle) from the LSAB+ Kit. Incubate for 15 minutes at room temperature.					
11. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.					
12. Apply the Label (red bottle) from the LSAB+ Kit. Incubate for 15 minutes at room temperature.					
13. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.					
(Add 1 drop of Da	AB per ml of substrate)	rk for 6 minutes at room temperature.  New Kit: yes / no			
15. Rinse the slides in	tap water 3 minutes.				
16. Counterstain with hematoxylin for 20 seconds.					
17. Rinse the slides in tap water until water is clear.					
18. Gently agitate slides in 1X wash buffer until the tissues turn blue.					
19. Dehydrate through the following solutions:					

95% Ethanol	1 time	3 minutes
100% Ethanol	3 times	3 minutes
Xylene	2 times	5 minutes

20. Coverslip

Updated 10/03/11